

Increased Resistance of Fluoroquinolones among *H. pylori* isolates from Patients with Gastroduodenal Diseases in Jordan

LUAY ABU-QATOUSEH, EYAD MALLAH, MONA BUSTAMI, EMAD ALKHATEEB
Faculty of Pharmacy and Medical Sciences
University of Petra
Airport Road, Amman
JORDAN

Abstract: - Recently, *Helicobacter pylori* has been connected to more than 80% of chronic active gastritis and other gastroduodenal diseases worldwide. Treatment of *H. pylori* is routinely dependent on the use of multiple antimicrobial agents however, recent data showed the emergence of resistance among clinical strains especially against metronidazole and clarithromycin. This study was conducted to investigate the rate of resistance to different antibiotics that are routinely used in the first line and second line therapies including ciprofloxacin and levofloxacin, tetracycline, amoxicillin, clarithromycin and metronidazole among *H. pylori* strains isolated from patients with gastroduodenal diseases in Jordan. Both antral and corpus mucosal biopsies from the stomach of patients with positive results of *H. pylori* stool antigen and urease breath tests were used for the isolation of *H. pylori* on selective culture media. The standard agar diffusion method was performed to determine the sensitivity of *H. pylori* clinical isolates against ciprofloxacin and levofloxacin according to CLSI. Among 62 *H. pylori* clinical strains isolated from gastric biopsies, 21% and 11% were resistant to levofloxacin and ciprofloxacin respectively. Resistance to metronidazole and clarithromycin was found in 90% and 11% respectively. No resistance was observed against amoxicillin, tetracycline and gemifloxacin. The following MIC₉₀ (mg/L) of resistant strains results were obtained at neutral pH 7.3, 64 for metronidazole, 2 for clarithromycin, 2 for ciprofloxacin and 1 for levofloxacin. The present study reported the emergence of increased resistance of fluoroquinolones among *H. pylori* clinical isolates in Jordan. Concern should be taken into consideration when triple and quadruple therapy regimens are applied for the management of *H. pylori* infections in our region

Key-Words: - *Helicobacter pylori*, Antibiotic resistance, fluoroquinolones, MIC, CLSI, Agar diffusion method

Received: February 7, 2020. Revised: May 31, 2020. Accepted: June 13, 2020. Published: June 24, 2020.

1 Introduction

Recently, interest in *Helicobacter pylori* related a disease is increasing. Several studies have been discussing the key role of *H. pylori* in the development of chronic gastritis and peptic ulcer worldwide [1, 2]. The ability of *H. pylori* to colonize the gastric mucosa constitutes a major risk factor in the pathogenesis of gastric cancer and gastric mucosa-associated lymphoid tissue (MALToma) [3-5]. *H. pylori* possess a variety of biological factors that enable this microaerophilic bacterium to withstand the extreme harsh conditions of human stomach [6]. Consequently, the eradication of *H. pylori* would primarily contribute to improve the clinical conditions of the patients infected with this bacterium including accelerating peptic ulcer healing and minimizing the recurrence of gastric cancer. In general, the current approaches for the treatment of diseases caused by *H. pylori*

rely basically on the effective eradication of this microorganism by the aid of at least two antibiotics and a proton pump inhibitor (triple and more recently quadruple therapy) [7]. This is achieved routinely by amoxicillin and clarithromycin in combination with a proton pump inhibitor given for a week [8]. Other treatments include a four-drug combination consisting of a PPI combined with bismuth (120 mg) and tetracycline (500 mg) four times daily and metronidazole (500 mg) three times daily [9]. However, due to the emergence of antibiotic resistance among *H. pylori* clinical strains particularly against metronidazole and clarithromycin, higher rates of failure to common regimens of were reported [10, 12]. Significantly, antimicrobial resistance patterns have not been uniformly reported [13]. Resistance rates reported vary from 10% to 90% for metronidazole, from 0% to 45% for clarithromycin, from 0% to 33% for

amoxicillin, from 5% to 59% for tetracycline, and from 6% to 21% for levofloxacin [14, 15, 16].

As there is an ongoing demand to determine resistance rates locally and to monitor these changing susceptibility patterns to recommend optimal therapy, this study is conducted to determine the prevalence of *H. pylori* resistance to various antibiotics especially those recently increasingly used such as fluoroquinolones and tetracycline of *H. pylori* isolates collected from patients with gastroduodenal diseases in Jordan.

2 Methods

2.1 Biopsy specimens and isolation of *H. pylori*

Antral and corpus mucosal gastric biopsies from stomach of patients with *H. pylori* related clinical manifestations including manifestations active gastritis, peptic ulcer or gastric cancer and positive results of stool antigen of *H. pylori* and urease breath test were obtained from the pathology department of Alpha medical diagnostic laboratories in Jordan. Gastric biopsy specimens were stored in cysteine Freezing medium at -80°C before processing in a sterile tissue grinder with heat-inactivated fetal bovine serum. Aliquots of 100 µl of the homogenate of each sample were directly sub-cultured onto Columbia blood agar base containing 7 % laked horse blood and selective supplement containing trimethoprim, vancomycin and polymyxin B (Oxoid, UK). All of the plates were incubated at 37°C under microaerophilic conditions using CampyGen atmosphere generating system (Oxoid, UK) for 3-5 days. Growth of *H. pylori* was confirmed on the basis of positive catalase, oxidase, and urease reactions; typical uniform, small, translucent colonies; curved Gram-negative bacilli on Gram-stained smears; susceptibility to cephalothin (30mg); and resistance to nalidixic acid (30mg) and subsequently by standard PCR of 16S rDNA test [26]. *H. pylori* cultures were stored at -70°C in Trypticase soy broth (Oxoid, UK) containing 10% v/v fetal calf serum (PAA, Austria) and 15% glycerol.

2.1 Antimicrobial susceptibility testing

Minimal inhibitory concentrations (MICs) were measured by agar dilution methods according to the Clinical and Laboratory Standards Institute (CLSI) as described by Abu-Qatouseh et al., 2017). In brief, bacterial suspensions were prepared to the 2 McFarland's standard and inoculated onto antibiotic-containing Mueller Hinton agar supplemented with 7% laked sheep blood. The MIC of each antibiotic was determined after 72 h of incubation. *H. pylori* NCTC 11916 was used as control strain.

The susceptibility of the isolates was tested using the E-test method (Epsilon meter test; AB Biodisk, Solna, Sweden) as recommended by the British Society for antimicrobial Chemotherapy (BSAC). The breakpoints used to classify strains as susceptible or resistant according to MIC are listed in Table 1. The breakpoints for clarithromycin and quinolones were interpreted according to CLSI recommendations. The breakpoints for amoxicillin, metronidazole, and tetracycline were interpreted according to the BSAC recommendations (not published by CLSI) [17].

3 Results and Discussion

A total of 62 *H. pylori* isolates were collected from endoscopic specimens of patients with gastroduodenal diseases. MIC₅₀ and MIC₉₀ values for the seven antibacterial agents used in this study are shown in Table 1. Best in vitro antimicrobial activity against *H. pylori* isolates was observed for amoxicillin, tetracycline and gemifloxacin were no resistance was reported.

The highest resistance was observed in metronidazole where only 6 strains were sensitive. Clarithromycin resistance was reported in 10 strains (16%). For fluoroquinolones, increased resistance to levofloxacin compared to ciprofloxacin was significantly reported with 15 (24%) and 10 (16%) strains are phenotypically resistant respectively (Table 2).

Table1: Minimum inhibitory concentration for the antimicrobial agents tested against 62 *H. pylori* isolates recovered in this study

Antimicrobial agent	Inhibitory concentration (mg/L)		
	MIC ₅₀	MIC ₉₀	MIC range
Amoxicillin	0.03	0.125	0.008-0.25
Tetracyclin	0.25	1	0.008-1
Metronidazole	32	128	1-256
Clarithromycin	0.06	0.25	0.008-4
Ciprofloxacin	0.5	1	0.016-32
Levofloxacin	0.25	1	0.03-8
Gemifloxacin	0.125	0.5	0.008-0.5

Table2: Rates of resistance to conventional antimicrobial agents among *H. pylori* isolates recovered in this study

Antimicrobial agent	Resistance rate (%)	Breakpoints criteria (mg/L)	
		S	R
Amoxicillin	0.0	≤ 1	≥ 2
Tetracyclin	0.0	≤ 2	≥ 4
Metronidazole	90	≤ 4	≥ 8
Clarithromycin	16	≤ 0.25	≥ 1
Ciprofloxacin	16	≤ 1	≥ 2
Levofloxacin	24	≤ 1	≥ 2
Gemifloxacin	0.0	≤ 1	≥ 2

The present study provides important updates regarding the prevalence of drug-resistant *H. pylori* in Jordan. According to our investigation, it has been revealed that amoxicillin, tetracycline, and gemifloxacin still show excellent in vitro anti-*H. pylori* activity. Approximately nine tenth and one fourth of isolates of *H. pylori* were resistant to Metronidazole and clarithromycin respectively. While our updated resistance rates to quinolones show a notable increase of resistance rate reported for this region (24% and 16% for both levofloxacin and ciprofloxacin respectively), rates for clarithromycin and metronidazole have minor increase over the last decade. Hence, more frequent

failure of *H. pylori* eradication would be anticipated when metronidazole is used in first-line regimens in areas where metronidazole resistance is prevalent, such as Jordan and other countries [13, 18, 19]. Since there is an increasing yet fluctuating patterns of antimicrobial resistance of *H. pylori* isolates, susceptibility testing (either phenotypic or genotypic) of clinical isolates of *H. pylori* in Jordan prior to empirical treatment would be optimal in terms of achieving higher eradication rates and better results.

It is unfortunately common in Jordan not to perform antimicrobial susceptibility testing of *H. pylori* to guide initial therapy; physicians should note that metronidazole resistance has been consistently associated with extra-gastric infections caused by other pathogenic microorganisms as well as due to the misuse of this antimicrobial agent. In addition, since metronidazole resistance rates depend primarily on the overall metronidazole consumption rates, its future role in empiric *H. pylori* therapy will continue to evolve depending on changing susceptibility rates. In contrast, amoxicillin showed excellent in vitro activity, indicating that amoxicillin should still be the first choice for treatment of *H. pylori* in Jordan. Clarithromycin has increased resistance rates which makes its use in the empiric clarithromycin-based triple therapy cautious. This is in agreement with most of the reports from similar geographical region however, the reported MIC values of clarithromycin resistant *H. pylori* in our study were higher. [20, 21]. In a study conducted by PCR for the detection of point mutations associated with clarithromycin resistance, similar rates were reported in Jordan [22].

Interesting in this study is the observation of minimal in vitro resistance to older drugs such as tetracycline. In addition, our investigation revealed that newer generation fluoroquinolones such as gemifloxacin were superior to ciprofloxacin and levofloxacin against clinical *H. pylori* isolates as in agreement with a previous report [23]. Higher resistance rates were observed to levofloxacin although MIC values are still near the lower breakpoints of resistance. Furthermore, ciprofloxacin resistance in *H. pylori* is increasing with higher MIC values. Whether gemifloxacin provides better clinical outcomes than other fluoroquinolones through fewer iterative failures warrants further studies. Nevertheless, these drugs may potentially be used as salvage therapy for patients for whom the clarithromycin-based standard therapy fails.

In Jordan, current eradication regimens available for *H. pylori* include triple drug combinations comprising a proton pump inhibitor and two antibiotics, amoxicillin plus metronidazole, clarithromycin plus metronidazole, or clarithromycin plus amoxicillin, with eradication rates of 60%, 50%, and 75%, respectively. Testing of fluoroquinolones susceptibility is not routinely performed for antibiotic resistance among *H. pylori* strains however, our results are in agreement with data from other countries which reported higher rates for fluoroquinolones resistance among *H. pylori* clinical strains [24, 25]. Although it is not possible to link eradication rates to the primary resistance rates to individual agents alone, we do believe that potential for synergism between various agents is implicated. Previous studies have demonstrated in vitro synergy between various combinations of amoxicillin, clarithromycin, and metronidazole with a proton-pump inhibitor for antibiotic-resistant strains of *H. pylori* [26, 27]. The interactions of fluoroquinolones especially levofloxacin with a second antibiotic, especially for clarithromycin-resistant strains, showed good antimicrobial synergism for the combination of levofloxacin (instead of amoxicillin) and clarithromycin. However, this depends on the MIC values of the clarithromycin where MIC values >1 mg/L would not have any useful effect of such drug combination. Nevertheless, a mounting evidence for testing the genetic determinants of antimicrobial susceptibility of clarithromycin is favoured.

4 Conclusion

In this study, we conclude that the rates of resistance among *H. pylori* clinical strains in Jordan for Metronidazole and fluoroquinolones are dramatically increasing. Caution should be taken for physicians when using these conventional antimicrobial agents in the treatment of *H. pylori* since it is expected to have higher treatment failure rates due the emerging resistance in *H. pylori* toward fluoroquinolones and metronidazole. In addition, it is recommended to perform antimicrobial susceptibility testing for *H. pylori* prior to use triple and quadruple therapies to minimize the increasing rates of resistance to the antimicrobial agents used in the medical practice and to minimize the undesirable outcomes associated with failure of treatment associated with *H. pylori* infections.

Future work is necessary to determine the mechanisms of resistance for fluoroquinolones in *H. pylori* since this can help in controlling spread of

resistance between the clinical isolates especially if plasmids are expected to be the responsible factors for the resistance phenotypes and genotypes.

References:

- [1] M. Hayama, Y. Kawakami, Y. Kaneko, K. Sano, H. Ota. Helicobacter pylori infection increases cell kinetics in human gastric epithelial cells without adhering to proliferating cells. *J Cell Mol Med*, vol 9, 2005, pp. 746-747.
- [2] Raj, Priya, John F. Thompson, and Debra H. Pan. Helicobacter pylori serology testing is a useful diagnostic screening tool for symptomatic inner city children. *Acta Paediatrica*, vol 106, 2017, pp. 470-477.
- [3] T. Shimizu, T. Akamatsu, A. Sugiyama H. Ota. Helicobacter pylori and the surface mucous gel layer of the human stomach. *Helicobacter*, vol 1, 1996, pp. 207-218.
- [4] Lanas, Angel, and Francis KL Chan. "Peptic ulcer disease. *The Lancet*, vol 390, 2017, pp 613-624.
- [5] C.Y. Wu, K.N. Kuo, M.S. Wu, Y.J. Chen, C.B. Wang, J.T. Lin. Early Helicobacter pylori eradication decreases risk of gastric cancer in patients with peptic ulcer disease. *Gastroenterology*, vol 137, 2009, pp. 1641-1648.
- [6] L. M. Brown. Helicobacter pylori: epidemiology and routes of transmission. *Epidemiol Rev*, vol 22, 2000, pp. 283-297.
- [7] J.G Kusters, A. an Vliet, E. Kuipers. Pathogenesis of Helicobacter pylori Infection. *Clin Microbiol Rev*, vol 19, 2006, pp. 449-490.
- [8] Take, S., Mizuno, M., Ishiki, K., Kusumoto, C., Imada, T., Hamada, F & Okada, H. Risk of gastric cancer in the second decade of follow-up after Helicobacter pylori eradication. *Journal of Gastroenterology*, vol 55, 2020, pp281-288.
- [9] K. Wolle, A. Leodolter, P. Malfertheiner, et al. Antibiotic susceptibility of Helicobacter pylori in Germany: stable primary resistance from 1995 to 2000. *J Med Microbiol*, vol, 51, 2002, pp.705-709.
- [10] M. Osato, R. Reddy, S. Reddy et al. Pattern of primary resistance of Helicobacter pylori to metronidazole or clarithromycin in the United States. *Arch Intern Med*, vol 161, 2001, pp.1217-1220.
- [11] A. Pilotto, M. Rassa, G. Leandro, et al. Prevalence of Helicobacter pylori resistance

- to antibiotics in Northeast Italy: a multicentre study. GISU. Interdisciplinary Group for the Study of Ulcer. *Dig Liver Dis*, vol 32, 2000, pp. 763–768.
- [12] L. Laine, R. Hunt, H. El-Zimaity et al. Bismuth-based quadruple therapy using a single capsule of bismuth biscaltrate, metronidazole, and tetracycline given with omeprazole versus omeprazole, amoxicillin, and clarithromycin for eradication of *Helicobacter pylori* in duodenal ulcer patients: a prospective, randomized, multicentre, North American trial. *Am J Gastroenterol*, vol 98, 2003, pp.562-567.
- [13] L. Abu-Qatouseh, M. Abu-Sini, A. M. A Mayyas, R. Darwish, T. Aburjai, I. Sabri. Molecular characterization and antibiotic susceptibility profiles of *Helicobacter pylori* isolated from patients with Gastrodeudenal diseases in Jordan. *International arab Journal of antimicrobial agents*, vol 6, 2017, pp. 1-8.
- [14] J Crowe, Sheila E. *Helicobacter pylori* infection. *New England Journal of Medicine* vol 380, no. 12, 2019, pp 1158-1165.
- [15] T.S. Chen, J.C. Luo, F.Y. Chang. Prevalence of *Helicobacter pylori* infection in duodenal ulcer and gastro-duodenal ulcer diseases in Taiwan. *J Gastroenterol Hepatol*, vol 25, 2010, pp. 919-922
- [16] H.Y. Lin, C.K. Chuang, H.C. Lee, N.C. Chiu, S.P. Lin, C.Y. Yeung A .seroepidemiologic study of *Helicobacter pylori* and hepatitis A virus infection in primary school students in Taipei. *J Microbiol Immunol Infect*, vol 38, 2005, pp. 176-182
- [17] Clinical and Laboratory Standards Institute Performance standards for antimicrobial susceptibility testing CLSI (2010) M100–S20, Table 2L M07.
- [18] H. Loghmani, F. Bdioui, W. Bouhlel, W. Melki, O. Hellara, N. Ben Chaabane, et al. Clarithromycin versus metronidazole in first-line *Helicobacter pylori* eradication. Prospective randomized study of 85 Tunisian adults. *Tunis Med*, vol 90, 2012, pp. 31-35.
- [19] W.L. Chang, B.S. Sheu, H.C. Cheng, Y.J. Yang, H.B. Yang, J.J. Wu Resistance to metronidazole, clarithromycin and levofloxacin of *Helicobacter pylori* before and after clarithromycin-based therapy in Taiwan *J Gastroenterol Hepatol*, vol 24, 2009, pp. 1230-1235.
- [20] N. Buta, N. Tanih, R. Ndip. Increasing trend of metronidazole resistance in the treatment of *Helicobacter pylori* infection: A global challenge. *Afr J Biotechnol*, 9(2010), pp. 1115-1121.
- [21] AT Abadi, T. Taghvaei, AM. Mobarez, BM. Carpenter, DS. Merrell. Frequency of antibiotic resistance in *Helicobacter pylori* strains isolated from the northern population of Iran. *J Microbiol*, vol 49, 2011, pp. 987-993.
- [22] A.F. Diab, DF. Hasan, SS. Nassar. Prevalence of *Helicobacter pylori* resistance to clarithromycin determined by 23S ribosomal RNA analysis in Jordan. *International arab journal of antimicrobial agents*, vol 6, 2016, pp. 1-5.
- [23] K.H. Hung, B.S. Sheu, W.L. Chang, H.M. Wu, C.C. Liu, J.J. Wu Prevalence of primary fluoroquinolone resistance among clinical isolates of *Helicobacter pylori* at a university hospital in Southern Taiwan. *Helicobacter*, vol 14, 2009, pp. 61-65.
- [24] J.M. Liou, C.Y. Chang, W.H. Sheng, Y.C. Wang, M.J. Chen, Y.C. Lee, et al. Genotypic resistance in *Helicobacter pylori* strains correlates with susceptibility test and treatment outcomes after levofloxacin- and clarithromycin-based therapies. *Antimicrob Agents Chemother*, vol 55, 2011, pp. 1123-1129.
- [25] A. Cuadrado-Lavin, J.R. Salcines-Caviedes, M.F. Carrascosa, P. Mellado, I. Monteagudo, J. Llorca, et al. Antimicrobial susceptibility of *Helicobacter pylori* to six antibiotics currently used in Spain. *J Antimicrob Chemother*, vol 67, 2012, pp. 170-173.
- [26] High prevalence of *Helicobacter pylori* infection with dual resistance to metronidazole and clarithromycin in Hong Kong. *Aliment Pharmacol Ther*, vol 14, 2000, pp. 901-910.
- [27] Verma, Anurag, Juhi Dubey, Rahul Rama Hegde, Vaibhav Rastogi, and J. K. Pandit. *Helicobacter pylori*: past, current and future treatment strategies with gastroretentive drug delivery systems. *Journal of drug targeting*, vol 24, 2016, pp. 897-915.

Contribution of individual authors to the creation of a scientific article (ghostwriting policy)

Luay Abu-Qatouseh, is the main author responsible for writing the manuscript.

Mona Bustami and Eyad Mallah, carried out data collection and statistical analysis.

Luay Abu-Qatouseh and Emad Al-khateeb have organized and executed the experiments.

Sources of funding for research presented in a scientific article or scientific article itself

This study was supported by the deanship of scientific research of university of Petra (11/4/2020)

Creative Commons Attribution License 4.0 (Attribution 4.0 International , CC BY 4.0)

This article is published under the terms of the Creative Commons Attribution License 4.0

https://creativecommons.org/licenses/by/4.0/deed.en_US